

<b>ALLIANCE FOR CLINICAL TRIALS IN ONCOLOGY</b>	<b>CORRELATIVE SCIENCE PROCEDURE MANUAL</b> Biospecimen Collection for A randomized phase II study of CHO(E)P vs. CC-486-CHO(E)P vs. duvelisib-CHO(E)P in previously untreated CD30 negative peripheral T-cell lymphomas Short Title- A051902	Version No: 2.4	Effective Date: 03/01/2024
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## CORRELATIVE SCIENCE PROCEDURE MANUAL

### 1. Purpose

This document describes the procedures required for the collection, shipping, and processing of biospecimens that are being submitted for patients enrolled or registered on A051902. This document also describes the procedures that will be followed subsequent to the receipt of biospecimens by the Alliance Biorepository at Washington University in St. Louis (ABWUSTL) prior to their use for protocol-specified and future, unspecified correlative science research studies. This document should be used by staff involved with any aspect of the A051902 biospecimen collection, processing, and submission, including staff at satellite institutions.

### 2. Scope

This document applies to all biospecimens collected specifically for the A051902 clinical trial study. Please refer to the trial protocol-specific language for additional details regarding eligibility, participant enrollment, and data submission. **Please ensure that you are reading the most updated version of this document. This document may experience minor updates, revisions, and clarifications independent of a formal protocol amendment. The most recent version of this document may be found on the Alliance website and CTSU.**

### 3. Definitions

Term	Definition
ABWUSTL	Alliance Biorepository at Washington University in St. Louis
FFPE	Formalin Fixed, Paraffin Embedded
IHC	Immunohistochemistry

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#### 4. Contact Information

<b>Protocol-related questions may be directed as follows:</b>	
<b>Questions</b>	<b>Contact (via email)</b>
Questions regarding patient eligibility, treatment, and dose modification:	Study Chair: Neha Mehta-Shah, MD, MSCI <a href="mailto:mehta-n@wustl.edu">mehta-n@wustl.edu</a> Nursing Contact: Maribeth Hohenstein, RN <a href="mailto:mahohens@unmc.edu">mahohens@unmc.edu</a> Protocol Coordinator: Destin Carlisle <a href="mailto:dcarlisle@bsd.uchicago.edu">dcarlisle@bsd.uchicago.edu</a> (where applicable) Data Manager: Andrea Kordaris-Corkill <a href="mailto:kordaris-corkill.andrea@mayo.edu">kordaris-corkill.andrea@mayo.edu</a>
Questions related to data submission, RAVE or patient follow-up:	Data Manager: Andrea Kordaris-Corkill <a href="mailto:kordaris-corkill.andrea@mayo.edu">kordaris-corkill.andrea@mayo.edu</a>
Questions regarding the protocol document and model informed consent:	Protocol Coordinator: Destin Carlisle <a href="mailto:dcarlisle@bsd.uchicago.edu">dcarlisle@bsd.uchicago.edu</a>
Questions related to IRB review	Alliance Regulatory Inbox <a href="mailto:regulatory@allianceNCTN.org">regulatory@allianceNCTN.org</a>
Questions regarding CTEP-AERS reporting:	Alliance Pharmacovigilance Inbox <a href="mailto:pharmacovigilance@alliancencn.org">pharmacovigilance@alliancencn.org</a>
Questions regarding specimens/specimen submissions:	Alliance Biorepository at Washington University (ABWUSTL) <a href="mailto:alliance@email.wustl.edu">alliance@email.wustl.edu</a>
Questions regarding drug supply	McKesson Clinical Research Services
Questions regarding drug administration	Pharmacy Contact: Barbara Todaro, PharmD <a href="mailto:barbara.todaro@roswellpark.org">barbara.todaro@roswellpark.org</a>

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- 4.1** For information on using the BiOMS system, please refer to the ‘Help’ links on the BiOMS webpage to access the online user manual, FAQs, and training videos. To report technical problems, such as login issues or application errors, please contact: 1-855-55-BIOMS or [bioms@alliancenctn.org](mailto:bioms@alliancenctn.org). For assistance in using the application or questions or problems related to specific specimen logging, please contact: 1-855-55-BIOMS or [bioms@alliancenctn.org](mailto:bioms@alliancenctn.org).
- 4.2** For all other questions regarding biospecimen procurement and shipping procedures, please contact the Alliance Biorepository Program Manager: 1-314-747-4402 or [alliance@email.wustl.edu](mailto:alliance@email.wustl.edu).

## 5. Site Preparation

- 5.1** Please refer to the A051902 protocol document for any specific requirements related to patient enrollment, registration, and regulatory compliance.
- 5.2** Please ensure that you have appropriate log on credentials and can successfully access the BiOMS application. The BiOMS application is used for logging the collection and shipment of biospecimens to the Alliance Biorepository at Washington University. For training and assistance in using the application or questions or problems related to specific specimen logging, please contact: 1-855-55-BIOMS or [bioms@alliancenctn.org](mailto:bioms@alliancenctn.org).

## 6. Collection Schema

The following biospecimens are to be collected at each of the time points listed in the table below. Please refer to biospecimen collection and processing methods and specific shipping procedures that are detailed in this manual.

**NOTE: NO SPECIMENS WILL BE SUBMITTED FOR PATIENTS ENROLLED TO THE SAFETY LEAD-IN COMPONENT.**

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Time Point	Kit (Y/N)	Biospecimen	Quantity	Collection / Processing Method	Shipping	Notes
<b>For all patients who consent to biobanking:</b>						
Registration	N	Fixed tissue block <b>OR</b> Stained <b>AND</b> Unstained tissue slides (4-5 um) <b>OR</b> Unstained tissue slides (4-5 um) <b>OR</b> Scanned slide images	1 block <b>OR</b> 15 total slides <b>OR</b> 15 total images	Fixed tissue block (9.2)  Fixed tissue slides (4-5 um) (9.3)  Scanned slide images (9.4)	Ambient	1, 2
Registration	N	Unstained tissue slides (10 um) <b>AND</b> Fixed tissue cores (4 mm)	5 (10 um) unstained slides <b>AND</b> 2 (4 mm) fixed tissue cores	Fixed tissue slides (10 um) (9.5)  Fixed tissue cores (4 mm) (9.6)	Ambient	1, 3
<b>For the first 12 patients registered to each arm (Phase II Safety Check) who consent to biobanking:</b>						
Cycle 1 Day 1 (+/- 3 days)	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
Cycle 1 Day 8	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
Up to 3 days before Cycle 2 Day 1	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
End of treatment visit (+/- 7 days)	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
Every 12 weeks (+/- 14 days) for 1 year post treatment	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
<b>For all patients in the Phase II Expansion who consent to biobanking:</b>						
Cycle 1 Day 1 (+/- 3 days)	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
Up to 3 days before Cycle 2 Day 1	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
End of treatment visit (+/- 7 days)	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4
24 weeks (+/- 14 days) post treatment	Y	Whole blood ( Streck BCT)	2 x 10 ml	Plasma for cfDNA (10.1)	Ambient	1, 4

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**Notes:**

1. Collection is optional for patients but requires all sites offer to patients to consent. Please see protocol-specific consent documents.
2. A fixed tissue block **OR** submission of a total of fifteen (15) fixed tissue slides is requested. Submission of a block is strongly preferred. Slides for the markers of interest (H&E, CD3, CD4, CD20, CD21 or CD23, EBER, BCL6, PD1, CXCL13, ICOS, SAP, and CD10) plus unstained slides to a total of 15 slides are requested. For example, if submitting stained slides for 4 of the requested markers, a total of 15-4=11 unstained slides should be submitted. If stained slides are not available for submission, 15 unstained slides can be sent. **See additional details in sections 9.2 and 9.3.** Alternatively, scanned slide images of slides with the stains listed above may be submitted in .SVS file format. **See additional instructions in section 9.4.**
3. **For patients who do not submit fixed tissue blocks**, an additional set of five (5) fixed tissue slides **AND** two (2) fixed tissue cores are requested. **For patients who have submitted blocks, slides and cores will be obtained at the Biorepository from the fixed tissue block submitted.** See additional details in sections 9.5 and 9.6.
4. Peripheral blood collection for cycle 1 day 1 must be done prior to initiation of chemotherapy. It may be collected at any point following registration to the study until initiation of chemotherapy. All peripheral blood collection time points may be drawn within the window permitted for the corresponding clinic visits outlined in **section 5.0** of the study protocol.

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## 7. Biospecimen Collection Kits

- 7.1** To facilitate the proper collection and shipping of whole blood in Streck BCT tubes, biospecimen collection kits and materials will be provided. The cost of the kit and shipping of the kit to the site will be paid for. The institution is expected to pay for shipping of the kit with the biospecimens back to the Alliance Biorepository at Washington University in St. Louis via priority overnight shipping.
- 7.2** Kits should be requested at least 10 working days in advance of the anticipated collection date. As many as 2 kits can be requested at one time. Since the collection materials (vacutainer tubes) have expiration dates, do not request kits more than 90 days prior to their anticipated use. All kits must be requested by using the BioMS system.
- 7.2.1** NOTE: Kits will be sent via FedEx at no additional cost to the participating institutions. Kits will not be sent via rush delivery service unless the participating institution provides their own FedEx® account number or alternate billing number for express service. The study will not cover the cost for rush delivery of kits.
- 7.2.2** Distributed kits will have a minimum shelf life of 90 days; unless precluded by the stability of a particular component.
- 7.3** Kit contents and specific instructions for use of the kit are provided in the kit box.
- 7.4** Once a kit is received, do not discard the outer cardboard overwrap. The kit, containing biospecimens, is to be shipped back in the same box.
- 7.5** Please return all components of the kit, regardless of whether they have been used or not. Kits and kit components are recycled when possible, minimizing the kit cost.
- 7.6** Well in advance of collecting biospecimens, inspect the biospecimen collection kit to ensure that all components are present and not expired, particularly if the kit has been onsite for longer than 30 days.
- 7.7** Note that individual kit components that are expired, damaged, or missing cannot be replaced. The remedy is to order a complete, new kit. Please note in your request that you are replacing an expired or damaged kit.

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- 7.8** Please return all kits that have expired or missing components. Return the ENTIRE kit using the cheapest possible shipping method at your expense. DO NOT DISCARD kits that have missing or expired components. Recycling kits keeps the cost of kit materials to a minimum. Please note that all out-going and incoming kits are tracked, and sites that have requested many more kits than they have returned will be charged for non-returned kits.
- 7.9** If a biospecimen collection component (e.g. vacutainer collection tube) is missing, damaged, or expired, the institution may substitute a like-kind collection tube from their own supply, if available.
- 7.10** Note that protocol requirements are based on blood volumes, not tube sizes. If the protocol requires the collection of 8 ml of whole blood, generally a 10 ml tube is provided in the kit for convenience. If desirable or necessary to collect 8 ml in 3 x 3 ml tubes (for example), that is permissible.
- 7.11** Because paraffin blocks or slides cut from such blocks may be requisitioned and received from the surgical pathology department at a different time than the day of procurement for other biospecimens, paraffin blocks, cores or cut slides may be sent independently of other biospecimens using the following guidelines:
- 7.11.1** There is no independent “kit” for submission of paraffin blocks, cores, slides, or whole slide image scans.
  - 7.11.2** Blocks and slides should be packaged to avoid breakage using a padded envelope or, preferably, a small Styrofoam container.
  - 7.11.3** During warm weather months, paraffin block, cores, and slides should be shipped in an insulated container that contains a refrigerant pack, to avoid heat > 25 degrees C (77 degrees F) that may melt paraffin and damage the tissue specimens.
  - 7.11.4** Blocks and slides may be shipped for standard overnight delivery according to institutional policies and using the preferred vendor.
- 7.12** Please see **Section 11 – Biospecimen Shipping** for specific instructions on shipping to ABWUSTL.

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## 8. Biospecimen Labeling and Tracking

- 8.1** All research biospecimens, with exception of surgical pathology blocks and tissue slides, **MUST** be labeled with the Alliance study number (A051902), Alliance patient ID number, patient initials (Last, First, Middle), the date and time (if applicable) of collection and specimen type. Specimen labels must match BioMS records.
- 8.2** Surgical pathology tissue blocks should be labeled according to institutional standards, including the surgical pathology number (e.g. "S16-1234") and the individual block identifier (e.g. "A3"). **Adhesive labels should not be used to cover the institutional label.** If tissue section slides are being submitted instead of the block, each tissue section slide should be labeled with the procurement date, Alliance study number (A051902), Alliance patient ID number, patient initials (Last, First, Middle), surgical pathology number (SPN) and institutional block ID, slide serial section number (1, 2, 3, etc.), and section thickness. Stained slides should additionally be labeled to indicate stain type (i.e. H&E, BCL6, CD10, BCER, etc.). **See section 9.4 for specific labeling instructions for WSI.**
- 8.3** **A de-identified copy of the surgical pathology report**, labeled with the Alliance patient ID number, is required to accompany **all** tissue submissions, including submission of WSI. Usually, this is generated by obscuring all PHI (names and dates) with white-out or a black magic marker, labeling each page of the report with the Alliance patient ID number, and photocopying the report. However, please make sure to **maintain the pathology accession numbers** so the submitted slides can be matched directly to the pathology report.
- 8.4** Label all containers and vials with an indelible, solvent-resistant marker when they are at ambient temperature.
- 8.5** Do not affix any labels to vials, slides or tubes. Label the collection containers directly with the marking pen.



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**8.6** All biospecimens that are collected and sent to ABWUSTL must be **logged and tracked in BioMS** under A051902. Do not send samples to the Biorepository in which the patient has not been registered onto the study. The BioMS system is a web-based application that tracks the collection and shipping of biospecimens. Once individual biospecimens are logged and ‘shipped’ in the BioMS system, a packing manifest will be created by the system. **This manifest must be printed out and must accompany all biospecimen shipments.** To become familiar with the BioMS system and for further information about training, access, and use, please contact the BioMS Help desk at: 1-855-55-BIOMS or [bioms@alliancenctn.org](mailto:bioms@alliancenctn.org).

**8.7** In the event that BioMS cannot be accessed, please complete a BioMS Specimen Log and Shipping Manifest form which can be found here- <http://tinyurl.com/alliance-biomscontingency>.

## 9. Tissue Collection

### 9.1 Overview.

**9.1.1** Please refer to protocol-specific instructions for procedures related to actual tissue procurement from individual participants. The method for research tissue procurement (needle core biopsy, sampling of surgically resected tumor) is dependent upon the disease site and the individual patient.

**9.1.2** When procuring tissue biospecimens by any method, when possible, avoid tissue that is grossly necrotic, hemorrhagic, fatty, or fibrous. If in doubt, briefly (1 min or less) place the tissue segment in a sterile specimen cup containing physiologic (normal) saline to rinse the tissue. Necrotic, hemorrhagic, and fatty tissue will generally dissolve or float on the surface while tumor and parenchymal tissue will remain intact and sink to the bottom of the cup.

### 9.2 Fixed Tissue Block

**9.2.1** For patients who consent to biobanking, a paraffin block from primary disease should be submitted.

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- 9.2.2** Any clinical surgical pathology block that is submitted for research studies will not be exhausted or rendered unsuitable for future diagnostic use. Any clinical surgical pathology block that is submitted will be returned within ten working days of written request, when needed for clinical management or clinical trial enrollment for a specific patient. Otherwise, all blocks will be returned to the submitting institution when the trial and correlative science study end points have been met.
- 9.2.3** In the event that an institution will not release a tissue block, the institution may instead submit stained and unstained tissue slides (up to a total of 15).
- 9.2.4** **During warm weather months, paraffin blocks and slides should be shipped in an insulated container that contains a refrigerant pack, to avoid heat > 25 degrees C (77degrees F) that may melt paraffin and damage the tissue specimens.**

### **9.3 Fixed Tissue Slides, Stained and Unstained (4-5 um)**

- 9.3.1** In cases where institutions are unable or unwilling to submit the requested tissue block, a set of 15 slides may be sent as an alternative.
- 9.3.2** Stained slides for the markers of interest (**H&E, CD3, CD4, CD20, CD21 or CD23, EBER, BCL6, PD1, CXCL13, ICOS, SAP and CD10**) plus unstained slides to a total of 15 slides are requested. For example, if submitting stained slides for 4 of the requested markers, a total of 15-4=11 unstained slides should be submitted.
- 9.3.3** If stained slides are not available for submission, a set of 15 unstained slides can be sent. Please follow the procedures below for submitting unstained tissue slides. If your pathology department is unwilling or unable to follow these tissue-sectioning instructions, please consider submission of a tissue block, which can be cut at the Biorepository and returned to your institution at a later date.
- 9.3.4** Unstained slides should be prepared following instructions below.

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# of slides	Section thickness	Slide type	Sample Types	Purpose
15	4-5 microns	Positively Charged	Primary tumor	Central Pathology Review

- 9.3.5** Serial, tissue sections should be **cut fresh** from the appropriate formalin fixed, paraffin embedded tissue block.
- 9.3.6** Cut sections at 4-5 micron thickness as indicated onto positively-charged slides.
- 9.3.7** Ensure that each slide is labeled with the procurement date, Alliance study number (A051902), Alliance patient ID number, patient initials (Last, First, Middle), surgical pathology number (SPN) and institutional block ID, slide serial section number (1, 2, 3, etc.), and thickness (4-5 microns). Please make sure to maintain the pathology accession numbers so the submitted slides can be matched directly to the pathology report.
- 9.3.8** Do not label slides with adhesive labels. Write or print information on the textured surface of the slide with indelible, solvent-resistant ink.
- 9.3.9** No adhesives or other additives should be used in the water bath.
- 9.3.10** Mount only one tissue section per slide. Make certain sections are placed on the painted / textured side of the slide.
- 9.3.11** When placing the sections onto the slides, ensure that the tissue is placed on the bottom third of the slide. Ensure that each serial section from the block is placed in the same orientation on each slide.
- 9.3.12** See figure below for proper mounting and labeling.

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03-09-2020 A051902	Procurement Date Alliance Study #
9110000 K, M, L S16-1234 B2	Alliance Patient ID # Patient Initials (L,F,M) SPN & Block ID
1 4 um	Serial Section # Thickness
	

**9.3.13** Air dry slides for 12-24 hours prior to shipping. Do not oven dry slides.

**9.3.14** During warm weather months, paraffin blocks and slides should be shipped in an insulated container that contains a refrigerant pack, to avoid heat > 25 degrees C (77degrees F) that may melt paraffin and damage the tissue specimens.

#### 9.4 Scanned Slide Images

- 9.4.1** In cases where institutions are unwilling or unable to submit the requested fixed tissue block or fixed tissue slides, scanned slide images of an H&E-stained slide and/or any other stained slides produced for histopathologic diagnosis required by the protocol may be submitted.
- 9.4.2** Scanned slide images will be uploaded via the Digital Pathology Image Portal (DPIP) in BioMS. Please see Appendix 1 for instructions on how to upload scanned slide images.
- 9.4.3** Unless otherwise specified in the protocol correlative science manual (CSM), scanned slide images must be created using an Aperio / Leica scanner at 40X magnification in an .SVS file format.
- 9.4.4** Scanned slide images should not contain PHI (patient name, date of birth, institutional surgical pathology number). The slide scanner should be configured to crop or exclude any image of the slide label. Alternatively, post-processing of the slide image file should remove the slide label image from the SVS file. **Images submitted with PHI or with the labels intact will not be accepted.**

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- 9.4.5** A digital de-identified pathology report (i.e. PDF file) must be included with all scanned slide image submissions, uploaded to BioMS. The surgical pathology number / accession number should be retained in order to correlate the report to the digital image specimen. The pathology report **must** also retain reference to individual block identifiers and must include the Alliance patient ID number and study ID so that each slide image can be correlated with the corresponding pathology report narrative. Please see Appendix 1 for instructions on how to upload the de-identified pathology report to BioMS.
- 9.4.6** The file name of the de-identified pathology report should correspond to the *Study ID, Alliance patient ID number, and last two digits of the institutional surgical pathology number* corresponding to the file name(s) assigned by DPIP.
- 9.4.7** Once the images have been uploaded, site staff should send an electronic copy of the manifest to [alliance@email.wustl.edu](mailto:alliance@email.wustl.edu).
- 9.4.8** Questions regarding appropriate image labeling, file naming conventions, and how to submit scanned slide images should be addressed to the Alliance Biorepository Protocol Management team ([alliance@email.wustl.edu](mailto:alliance@email.wustl.edu)). Questions related to BioMS should be addressed to the BioMS help desk ([bioms@alliancenctn.org](mailto:bioms@alliancenctn.org))

## 9.5 Fixed Tissue Slides, Unstained (10 um)

- 9.5.1** In cases where institutions have **NOT** submitted fixed tissue blocks, a set of five (5) unstained slides cut at 10 um are requested in addition to the 15 unstained/stained slides cut at 4-5 um (**section 9.3**). Two fixed tissue cores should also be submitted with the unstained slides (**see section 9.6**).
- 9.5.2** Please follow the procedures below for submitting unstained tissue slides. If your pathology department is unwilling or unable to follow these tissue-sectioning instructions, please consider submission of a tissue block, which can be cut at the Biorepository and returned to your institution at a later date.
- 9.5.3** Unstained slides should be prepared following instructions below.

# of slides	Section thickness	Slide type	Sample Types	Purpose
5	10 micron	Non-charged	Primary tumor	DNA/RNA

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**9.5.3.1** Tissue should be mounted and slides should be labeled following instructions in **section 9.3**.

**9.5.3.2** During warm weather months, paraffin blocks and slides should be shipped in an insulated container that contains a refrigerant pack, to avoid heat > 25 degrees C (77degrees F) that may melt paraffin and damage the tissue specimens.

## 9.6 Fixed Tissue Cores

**9.6.1** In cases where institutions have **NOT** submitted fixed tissue blocks, two fixed tissue cores are requested in addition to the 15 stained/unstained slides cut at 4-5 um and the 5 unstained slides cut at 10 um. Two (2) 4mm cores should be submitted from the primary tissue block.

**9.6.2** Place the tissue cores directly into microcentrifuge tubes (one core per tube) or any other suitable container. Label the tubes of tissue following the guidelines outlined in **section 8**.

## 10. Blood Collection Methods

### 10.1 Plasma Nucleic Acid (Streck) Tube Processing

**10.1.1** Collect 10 ml of blood into each of the Streck BCT tubes using standard venous phlebotomy. Invert tubes 10 times.

**10.1.2** Store Streck tubes with whole blood at room temperature. Do not freeze or refrigerate the tubes. The tubes may be stored for up to 72 hours at ambient temperature before shipment. Ensure that the Streck tubes are shipped at ambient temperature to avoid freezing.

**10.1.3** During warm weather months (i.e. June—August), a refrigerated pack (not frozen) should be included in the shipment to maintain ambient temperature. When shipping during other months of the year, a room temperature pack should be included in the shipment.

<b>ALLIANCE FOR CLINICAL TRIALS IN ONCOLOGY</b>	<b>CORRELATIVE SCIENCE PROCEDURE MANUAL</b> Biospecimen Collection for A randomized phase II study of CHO(E)P vs. CC-486-CHO(E)P vs. duvelisib-CHO(E)P in previously untreated CD30 negative peripheral T-cell lymphomas Short Title- A051902	Version No: 2.4	Effective Date: 03/01/2024
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## 11. Biospecimen Shipping

### 11.1 Overview

- 11.1.1** Please see the Directions for Use (DFU) document that is included in each kit for specific directions on how to package and ship biospecimens.
- 11.1.2** Place the original, completed copy of the BioMS packing manifest in the shipment. If sending tissue or scanned slide images, include a copy of the de-identified surgical pathology report. Do not send specimens without a completed BioMS Packing Manifest or substitute "BioMS Downtime Form." Biospecimens cannot be accepted without this completed form.
- 11.1.3** All biospecimens should be shipped on Monday – Thursday only. If physician or patient schedules mandate a Friday, weekend, or holiday collection, please contact the Alliance Biorepository Program Manager: 1-314-747-4402 or [alliance@email.wustl.edu](mailto:alliance@email.wustl.edu) for further instructions, at least 24 hours prior to anticipated collection.
- 11.1.4** Do not ship on Friday, Saturday, Sunday or the day before a nationally recognized holiday.

### 11.2 Shipping to ABWUSTL

- 11.2.1** Ship container for PRIORITY OVERNIGHT DELIVERY according to IATA guidelines and standard institutional policies and using the preferred vendor. Ship specimens to ABWUSTL on Monday through Thursday only. Do not ship on Fridays, Saturdays or on the day before a national holiday.

Ship to:

**Alliance Biorepository at Washington University in St. Louis**  
**c/o Siteman Cancer Center Tissue Procurement Core**  
**Washington Univ. School of Medicine**  
**425 S. Euclid Ave.**  
**Room 5120**  
**St. Louis, MO**  
**63110-1005**  
**Phone: 314-454-7615**

<b>ALLIANCE FOR CLINICAL TRIALS IN ONCOLOGY</b>	<b>CORRELATIVE SCIENCE PROCEDURE MANUAL</b> Biospecimen Collection for A randomized phase II study of CHO(E)P vs. CC-486-CHO(E)P vs. duvelisib-CHO(E)P in previously untreated CD30 negative peripheral T-cell lymphomas Short Title- A051902	Version No: 2.4	Effective Date: 03/01/2024
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## 12. ABWUSTL Biospecimen Receipt and Quality Assurance Measures

- 12.1** Upon receipt, all biospecimens will be accessioned into the TPC informatics system, OpenSpecimen.
- 12.2** All biospecimens will be logged, associated, and tracked by the unique patient biopsy control number.
- 12.3** Each individual biospecimen will receive and be physically labeled with a unique biospecimen identifier, associated with the biopsy control number in the TPC informatics system.
- 12.4** Upon receipt, all physical biospecimens received will be reconciled with what is recorded on the BioMS packing manifest. Any discrepancies noted will be communicated to the Program Manager who will contact the submitting site for reconciliation.
- 12.5** Upon receipt, any biospecimen received that is not in appropriate physical condition (broken vials, frozen samples that are thawed, ambient samples that are frozen) will be reported to the Program Manager, who will contact the submitting site for reconciliation.
- 12.6** Aliquoted biofluids will be stored under liquid nitrogen vapor.
- 12.7** Fixed tissue blocks will be vacuum packed to prevent oxidation and stored at 4 degrees Celsius (refrigerated) to minimize degradation of cellular antigens.
- 12.8** All biospecimens will remain in storage until additional processing or review is requested in writing by the appropriate protocol PI.

## 13. Document History

Version	Description and Justification of Change	Author	Effective Date
2.4	Added additional stain to send if sending slides for registration tissue.	KL	03/01/2024
2.3	Changed instructions for scanned slide image submission	KL	02/01/2024
2.2	Added new instructions for WSI submission	KL	10/01/2023
2.1	Changed instructions for WSI submissions, added warm weather shipping instructions, added contact table	KL	06/13/2023
2.0	Add instructions for WSI scan submissions	AAW	03/21/2023
1.1	Clarify collection for Phase II safety check	PAA	05/12/2021
1.0	New	PAA	03/30/2021

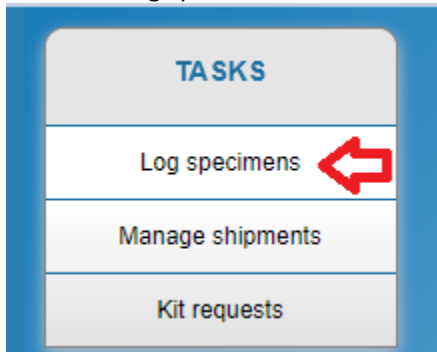


<b>ALLIANCE FOR CLINICAL TRIALS IN ONCOLOGY</b>	<b>CORRELATIVE SCIENCE PROCEDURE MANUAL</b> Biospecimen Collection for A randomized phase II study of CHO(E)P vs. CC-486-CHO(E)P vs. duvelisib-CHO(E)P in previously untreated CD30 negative peripheral T-cell lymphomas Short Title- A051902	Version No: 2.4	Effective Date: 03/01/2024
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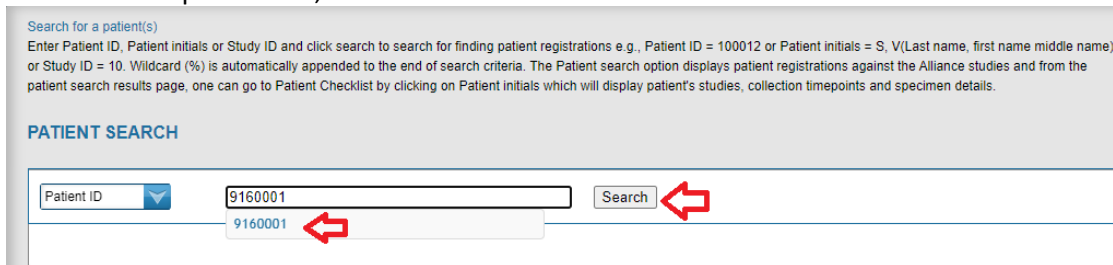
Appendix 1 – Digital Pathology Image Portal Instructions

Please note that while the Study ID and specimen requirement title might be different than your study, the concept is still the same.

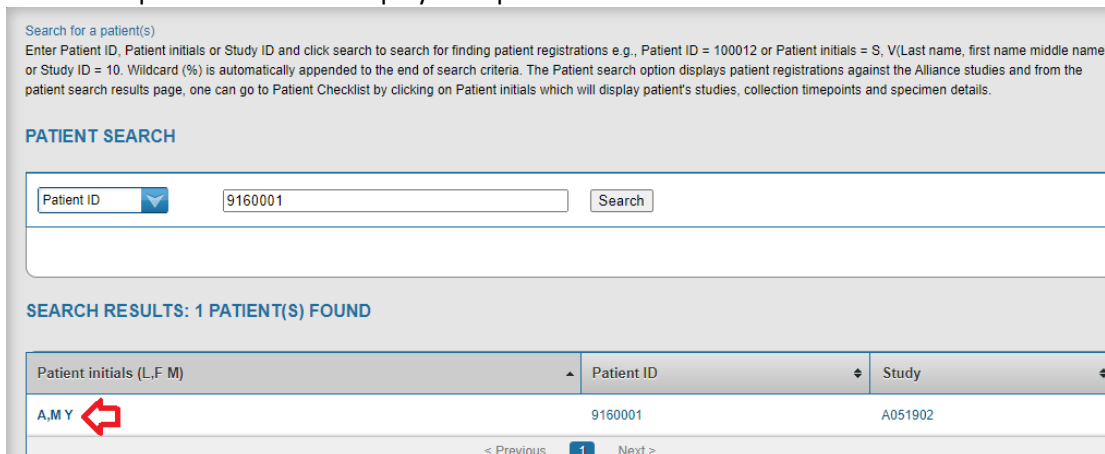
1. Login BioMS <https://bioms.wustl.edu/bioms/login>
2. Select "Log specimens" from the Task menu.



3. Enter the patient ID (obtained when registering the patient in OPEN), in the search field, select the matching ID from the drop-down list, click "Search."



4. Click the patient initials to display the Specimen Checklist view.



5. Click on the Alliance Digital Pathology Image (DPI) Portal Icon. Do not enter the slide count or collection date as only specimens in a "Pending" status can have images uploaded.

Collected	Study	Specimen expected	Quantity	Collection date	Status	Actions
<b>Consented to Biobanking : Registration</b>						
<input type="checkbox"/>	A051902	FFPE Block- Tumor	1.0 count	<input type="text"/>	14 Not Collected	
<input type="checkbox"/>	A051902	Unstained Slides (4-5um)	0.0 count	<input type="text"/>	14 Pending	
<input type="checkbox"/>	A051902	5 Unstained Slides (10um)- Tumor	5.0 count	<input type="text"/>	14 Pending	
<input type="checkbox"/>	A051902	Stained Slides (4-5um)	0.0 count	<input type="text"/>	14 Pending	
<input type="checkbox"/>	A051902	2 Fixed Tissue Cores (4mm)- Tumor	2.0 gm	<input type="text"/>	14 Pending	
<input type="checkbox"/>	A051902	Scanned Slide Images	1.0 count	<input type="text"/>	14 Pending	

Save Go to shipments

6. A new browser tab will open displaying the DPI Portal. This tab must remain open during the entire loading process. Closing the new browser will terminate the loading process.

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*) and click on 'Choose File' to select a file. To add more files, click on the 'Add' button. Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

<b>Study ID:</b> A051902	<b>Patient ID:</b> 9160001	<b>Collection Arm:</b> Consented to Biobanking	<b>Collection Event:</b> Registration
-----------------------------	-------------------------------	---	--

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *	Upload File name	Upload progress
<input type="text"/>	<input type="text"/>	Please select	Choose File No file chosen		

Add Upload File Cancel

7. Please note the instructions at the top of the page. All files must be in .SVS format and cannot exceed 5GB  
 8. Enter the last 2 digits of the Surgical Pathology Number. (Example if the SPN is 24-12345, then enter 45)

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*) and click on 'Choose File' to select a file. To add more files, click on the 'Add' button. Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

<b>Study ID:</b> A051902	<b>Patient ID:</b> 9160001	<b>Collection Arm:</b> Consented to Biobanking	<b>Collection Event:</b> Registration
-----------------------------	-------------------------------	---	--

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
45	<input type="text"/>	Please select	Choose File No file chosen

Add Upload File Cancel

9. Enter the block ID (Example: if the slide was cut from block A2 then scanned, enter A2)

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*). Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. You can continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the naming convention in the window before the transfer is complete, you will need to upload the files again.

**Study ID:** A051902      **Patient ID:** 9160001      **Collection Arm:** Consented to Biobanking      **Collection Event:** Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
<input type="text" value="45"/>	<input type="text" value="A2"/>	Please select	<input type="button" value="Choose File"/> No file chosen

10. Select the "Stain Type" from the drop-down list.

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*). Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. You can continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the naming convention in the window before the transfer is complete, you will need to upload the files again.

**Study ID:** A051902      **Patient ID:** 9160001      **Collection Arm:** Consented to Biobanking      **Collection Event:** Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
<input type="text" value="45"/>	<input type="text" value="A2"/>	Please select	<input type="button" value="Choose File"/> No file chosen

11. Click Choose File, then select the file from your computer directory.

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*). Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. You can continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the naming convention in the window before the transfer is complete, you will need to upload the files again.

**Study ID:** A051902      **Patient ID:** 9160001      **Collection Arm:** Consented to Biobanking      **Collection Event:** Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
<input type="text" value="45"/>	<input type="text" value="A2"/>	Hematoxylin and Eosin (H&E)	<input type="button" value="Choose File"/> No file chosen

12. BioMS will then rename the file to the DPI format (StudyID\_PatientID\_Last2Digitsof SPN\_BlockID\_Stain Type\_.svs

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*) and click on 'Choose File' to select a file. To add more files, click on the 'Add' button. Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

Study ID: A051902      Patient ID: 9160001      Collection Arm: Consented to Biobanking      Collection Event: Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *	Upload File name	Upload progress
<input type="text" value="45"/>	<input type="text" value="A2"/>	Hematoxylin and Eosin (H&E) ▼	<input type="button" value="Choose File"/> patient_1_HE.svs	A051902_9160001_45_A2_HE.svs	

13. If there are additional files to load for the patient, click Add and repeat steps 8-11

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*). Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

Study ID: A051902      Patient ID: 9160001      Collection Arm: Consented to Biobanking      Collection Event: Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
<input type="text" value="45"/>	<input type="text" value="A2"/>	Hematoxylin and Eosin (H&E) ▼	<input type="button" value="Choose File"/> patient_1_HE.svs
<input type="text"/>	<input type="text"/>	Please select ▼	<input type="button" value="Choose File"/> No file chosen

14. Once all files are selected, click "Upload File."

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*). Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

Study ID: A051902      Patient ID: 9160001      Collection Arm: Consented to Biobanking      Collection Event: Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *
<input type="text" value="45"/>	<input type="text" value="A2"/>	Hematoxylin and Eosin (H&E) ▼	<input type="button" value="Choose File"/> patient_1_HE.svs
<input type="text" value="45"/>	<input type="text" value="A2"/>	Ki67 (K67) ▼	<input type="button" value="Choose File"/> patient_1_Ki67.svs

15. BioMS will display a progress bar for each file. DO NOT close the DPI window as it will terminate the load process. Users can navigate to other windows during the load process.

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*) and click on 'Choose File' to select a file. To add more files, click on the 'Add' button. Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

Study ID: A051902      Patient ID: 9160001      Collection Arm: Consented to Biobanking      Collection Event: Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *	Upload File name	Upload progress
45	A2	Hematoxylin and Eosin (H&E)	Choose File patient_1_HE.svs	A051902_9160001_45_A2_HE.svs	24%
45	A2	Ki67 (K67)	Choose File patient_1_Ki67.svs	A051902_9160001_45_A2_K67.svs	35%

Add      Upload File      Cancel

**Warning:** Please ensure that this window remains open after initiating a file transfer request. Once the file transfer is complete, you will receive a completion message on this page. At that time, you can exit and close this page. While the file transfer is in progress, feel free to continue working in BioMS or other tasks. However, please **DO NOT CLOSE THIS PAGE** until the transfer is finished.      Uploading

16. BioMS will display a successful message in the Alliance DPI portal and send an email to the user once all images are transferred.

**Alliance Digital Pathology Image (DPI) Portal**

**Instructions:** The file format must be .svs, and each one cannot exceed 5GB. You can submit up to 15 files at once. Complete all mandatory fields (marked by \*) and click on 'Choose File' to select a file. To add more files, click on the 'Add' button. Finally, click 'Upload File' to initiate transfer. Uploading may take time due to the large size of file(s), and the transfer time depends on your internet upload speed. File transfer will be processed asynchronously, allowing you to continue using BioMS or perform work on this PC. However, you must keep this window open and cannot close it. The file you upload will be renamed as per the Alliance DPI Standard Operating Procedures. If you happen to close this window before the transfer is complete, you will need to upload the files again.

Study ID: A051902      Patient ID: 9160001      Collection Arm: Consented to Biobanking      Collection Event: Registration

Last 2 digits of Surgical Pathology# *	Block ID: *	Stain Type *	Select file to upload *	Upload File name	Upload progress
45	A2	Hematoxylin and Eosin (H&E)	Choose File patient_1_HE.svs	A051902_9160001_45_A2_HE.svs	Completed
45	A2	Ki67 (K67)	Choose File patient_1_Ki67.svs	A051902_9160001_45_A2_K67.svs	Completed

Add      Upload File      Exit

**File Upload Completed Successfully!**  
Please click on the BioMS Patient Checklist link below to mark the Digital Specimen as collected and proceed to ship it to the respective repository. If you are not currently logged into BioMS, please log in on a different tab and then click on the BioMS Patient Checklist link below to complete the Digital Specimen collection and shipment process. Thank you.  
[BioMS Patient Checklist](#)

- 17. The Alliance DPI portal window may now be closed.
- 18. Navigate back to the patient's check list view to now log the images as "collected."
- 19. Enter the number of images loaded in the "Quantity" field.
- 20. Enter the date of the procedure (biopsy, surgery) in the "Collection Date."

Registration      On Therapy

Collected	Study	Specimen expected	Quantity	Collection date	Status	Actions
<b>Consented to Biobanking : Registration</b>						
<input type="checkbox"/>	A051902	FFPE Block- Tumor	1.0 count	14	Not Collected	
<input type="checkbox"/>	A051902	Unstained Slides (4-5um)	0.0 count	14	Pending	
<input type="checkbox"/>	A051902	5 Unstained Slides (10um)- Tumor	5.0 count	14	Pending	
<input type="checkbox"/>	A051902	Stained Slides (4-5um)	0.0 count	14	Pending	
<input type="checkbox"/>	A051902	2 Fixed Tissue Cores (4mm)- Tumor	2.0 gm	14	Pending	
<input checked="" type="checkbox"/>	A051902	Scanned Slide Images	2 count	10/19/2022 00:00	Pending	

21. Click the "Clip board" icon to load a digital copy of the corresponding surgical pathology report.

Registration		On Therapy				
Collected	Study	Specimen expected	Quantity	Collection date	Status	Actions
<b>Consented to Biobanking : Registration</b>						
<input type="checkbox"/>	A051902	FFPE Block- Tumor	1.0 count		14 Not Collected	
<input type="checkbox"/>	A051902	Unstained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	5 Unstained Slides (10um)- Tumor	5.0 count		14 Pending	
<input type="checkbox"/>	A051902	Stained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	2 Fixed Tissue Cores (4mm)- Tumor	2.0 gm		14 Pending	
<input checked="" type="checkbox"/>	A051902	Scanned Slide Images	2 count	10/19/2022 00:00	14 Pending	

**SPECIMEN COMMENTS**

**⚠ If you are submitting tissue from a pathology block, slides, curls, etc. PLEASE remember to include a printed, de-identified copy of the corresponding pathology report in your shipment.**

**SPECIMEN ATTACHMENTS**

Title	Attachment	Action
<input type="text" value="Path Report Patient 1"/>	<input type="button" value="Choose File"/> A051902	
<input type="button" value="Add more"/>		

22. Click Save

Collected	Study	Specimen expected	Quantity	Collection date	Status	Actions
<b>Consented to Biobanking : Registration</b>						
<input type="checkbox"/>	A051902	FFPE Block- Tumor	1.0 count		14 Not Collected	
<input type="checkbox"/>	A051902	Unstained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	5 Unstained Slides (10um)- Tumor	5.0 count		14 Pending	
<input type="checkbox"/>	A051902	Stained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	2 Fixed Tissue Cores (4mm)- Tumor	2.0 gm		14 Pending	
<input checked="" type="checkbox"/>	A051902	Scanned Slide Images	2.0 count	10/19/2022 00:00	14 Collected	

Save
 Go to shipments

23. Click "Go To Shipments" to now create the digital shipment

Collected	Study	Specimen expected	Quantity	Collection date	Status	Actions
<b>Consented to Biobanking : Registration</b>						
<input type="checkbox"/>	A051902	FFPE Block- Tumor	1.0 count		14 Not Collected	
<input type="checkbox"/>	A051902	Unstained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	5 Unstained Slides (10um)- Tumor	5.0 count		14 Pending	
<input type="checkbox"/>	A051902	Stained Slides (4-5um)	0.0 count		14 Pending	
<input type="checkbox"/>	A051902	2 Fixed Tissue Cores (4mm)- Tumor	2.0 gm		14 Pending	
<input checked="" type="checkbox"/>	A051902	Scanned Slide Images	2.0 count	10/19/2022 00:00	14 Collected	

Save
 Go to shipments

24. From the Manage Shipment page, check the box for the specimen requirement, leave the shipment default to "Auto" and click the green arrow pointing to the right.



**SPECIMENS AND SHIPMENTS**

Filter By Ship To:

**SPECIMENS FOR 9160001 (A,M Y) AND STUDY A051902**

All

<input type="checkbox"/>	Specimen ID & Title	Patient ID & Initials	Study & TimePoint
<input checked="" type="checkbox"/>	135090740 - Scanned Slide Images	9160001 (A,M,Y)	A051902 - Registration

Auto  
Manual

→

→

←

25. BioMS will move the specimen requirement into an appropriate shipment. The shipment ID containing the specimen requirement will be displayed at the top of the screen.

Specimen(s) added to shipment(s) [135141040]

Logged into BioMS as BRINKA Logout ? Help

Review Pending Specimens and Shipments

Review the list of specimens and shipments pending to be shipped. To view the contents of a particular shipment, click on downward arrow icon below the box icon for the shipment. Select the shipment by clicking on the round button above the box icon for the shipment, then click on the appropriate button on the top/bottom of the shipments section to perform a particular action for the selected shipment.

**SPECIMENS AND SHIPMENTS**

Filter By Ship To:

**SPECIMENS FOR 9160001 (A,M Y) AND STUDY A051902**

All

<input type="checkbox"/>	Specimen ID & Title	Patient ID & Initials	Study & TimePoint
<input type="checkbox"/>			

Auto  
Manual

→

←

**SHIPMENTS**

Create Send Delete Print Edit

<input type="radio"/>		Shipment ID: 135141040
		Ship to: Alliance Biorepository Washington University St. Louis
		Ship method: Unspecified
		Status: Not Shipped

26. Click the radio button for the shipment, then click Send.

**SHIPMENTS**

Create Send Delete Print Edit


Shipment ID:	135141040
Ship to:	Alliance Biorepository Washington University St. Louis
Ship method:	Unspecified
Status	Not Shipped

▼(1)


27. On the manifest page, enter your phone number, email address.
28. Select "Digital Upload" as the carrier.
29. Enter "N/A" for the tracking number.
30. Enter the date the images were transferred / uploaded as the shipment date.
31. Click Send Shipment.

Send shipment
← Back


**BioMS ShipmentID**



135141040

<b>Shipment ID</b>	135141040	<b>Status</b>	Not Shipped	<b>Carrier *</b>	Digital Upload
<b>Sender site *</b>	Washington University School of Medicine	<b>Receiver site</b>	Alliance Biorepository Washington University St. Louis	<b>Tracking number</b>	N/A
<b>From *</b>	660 South Euclid Avenue Saint Louis, Missouri 63110	<b>To</b>	425 S. Euclid Ave. RM 5120 St. Louis, Missouri 63101	<b>Shipping date *</b>	01/18/2024 
<b>Sender contact *</b>	Amy Brink	<b>Receiving site email</b>	tbank@wudosis.wustl.edu	<b>Received date</b>	
<b>Phone *</b>	999-999-9999	<b>Receiving site phone</b>	(314) 454-7615	<b>Notes</b>	
<b>Sender email *</b>	abrink@wustl.edu				

**CONTENTS**

BioMS Specimen ID	Specimen	Qty	Collection Date	Patient ID	Study ID	Epoch, Arm, CPE
 135090740	Scanned Slide Images	2.0 count	10/19/2022 00:00	9160001 (A,M Y)	A051902	Registration, Consented to Biobanking, Registration

32. Email a copy for the BioMS manifest to [alliance@email.wustl.edu](mailto:alliance@email.wustl.edu)

For additional information and screen shots please visit

<https://washu.atlassian.net/wiki/spaces/BP/pages/468844574/Alliance+Digital+Pathology+Image+Portal+DPI>